

# Loading your own images into Athena Zebrafish software

## IDEA Bio-Medical Seeing Better

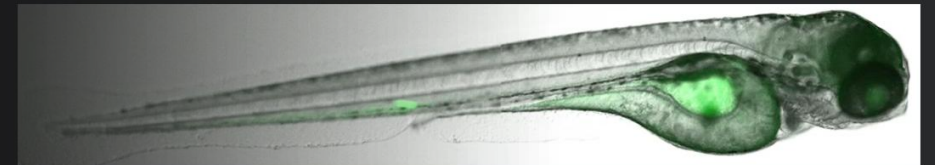
Please begin, first, with our quick-start guide to analyze the sample images included with the software download.

Access the quick-start guide [HERE](#).



### Zebrafish Analysis

Automatic quantification of Zebrafish embryo for studying of morphological features, fluorescence measurements and internal organelle properties



# Preparing Your Images for Analysis in Athena

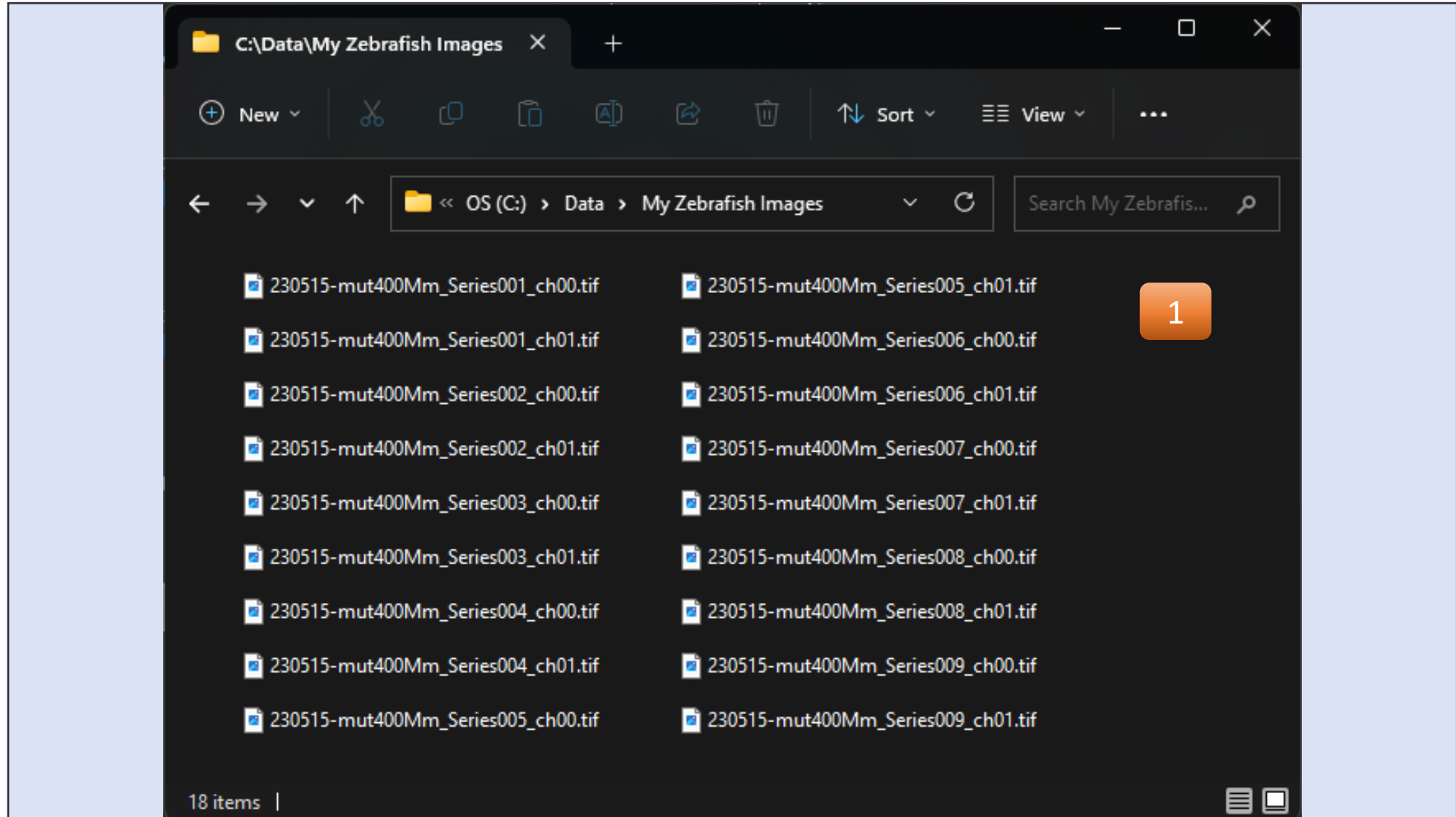
## *Two-channel tiff images example*

1. In this example, standard .tif images are saved into a folder on the local hard drive; the path is displayed in the windows explorer.

This dataset contains two color channels, each indicated in the filename with recognizable text:  
Brightfield = *ch01*  
Green fluor. = *ch00*

Other text in the filename, as shown, will not affect loading the images into Athena.

If images contain well-identifiers, such as A1, B2, C3, etc., then the images will be assigned to those wells.



# Open Athena

## *Shortcut from desktop*

2. Open Athena using the shortcut on your desktop.

Sample images that can be analyzed for free are also accessible on the desktop from the 'Samplefish' folder shortcut.



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Seeing Better

# Opening Athena

## *Selecting user & updating license*

3. Select the default 'Admin' user, or other user profile, from the menu.

If you need to insert a license code or coupon code, please see the quick-start guide by clicking [HERE](#).



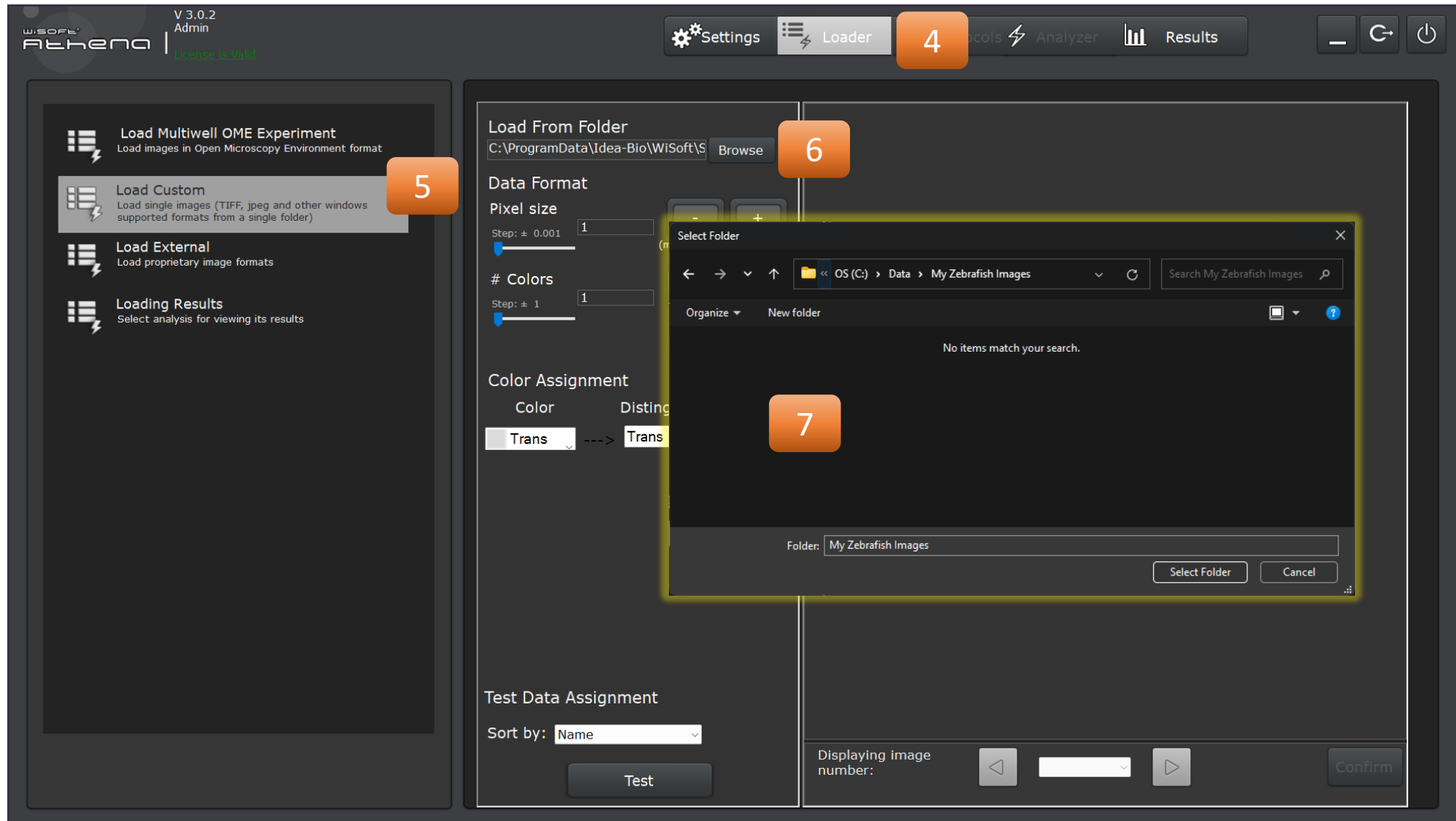
# Using Athena with Your Own Images

## *Define where images are located*

4. Select the 'Loader' tab to open images.
5. Select the 'Load Custom' option from the menu on the left.
6. Press the 'Browse' button to navigate to the **FOLDER** location where the tiff images you would like to load & analyze are located.
7. The file explorer window shows the currently selected folder. Navigate to your image storage location, then click the "Select Folder" button to choose this path.

### *Info:*

The default folder path is pre-set to identify the freely analyzable sample images. Other parameters are also set to permit loading the sample images.



# Using Athena with Your Own Images

## *Define characteristics of your images*

8. Define the correct pixel size for your images by clicking on the text field to open the number pad.
9. Use the numbers on screen or your keyboard to set the desired pixel size.

Click 'Done' to set the pixel size.

Using the default pixel size of 1 will return areas in dimensions of pixels, not microns.

### *Info:*

Contact your microscope manufacturer if you have questions about how to find the pixel size for your images. In some cases, metadata such as pixel size may sometimes be saved in an associated .xml file, so can be found therein.

The screenshot displays the Athena software interface (V 3.0.2 Admin) with the 'Loader' tab selected. The 'Load From Folder' section is active, showing the path 'C:\Data\My Zebrafish Images' and a 'Browse' button. Under 'Data Format', the 'Pixel size' is set to 1, and a numeric keypad overlay is visible with the number 8 highlighted. The '# Colors' is set to 1. The 'Color Assignment' section shows 'Color' set to 'Trans' and 'Distinguishing Text' set to 'Trans'. The 'Test Data Assignment' section shows 'Sort by' set to 'Name' and a 'Test' button. A numeric keypad overlay is also visible, showing the number 9 highlighted. The keypad has a display showing '3.196' and buttons for digits 1-9, 0, and a decimal point, along with 'Del', 'Cancel', and 'Done' buttons. An information icon is also present on the keypad.

# Using Athena with Your Own Images

## *Define characteristics of your images*

10. Define the number of colors present in your dataset.
11. Define the channel that should be assigned to each of the colors present.

### *Info:*

The “Distinguishing Text” editable fields are auto-filled with default values, as shown. These text strings are used to assign each image file in the selected data folder (steps 6 & 7) to one of the colors for analysis.

The screenshot displays the Athena software interface for loading and configuring images. The top navigation bar includes 'Settings', 'Loader', 'Protocols', 'Analyzer', and 'Results' tabs. The main interface is divided into three panels:

- Left Panel:** Contains loading options: 'Load Multiwell OME Experiment', 'Load Custom' (selected), 'Load External', and 'Loading Results'.
- Middle Panel:** Titled 'Load From Folder', it includes:
  - Data Format:** 'Pixel size' set to 3.196 (mic) with step ± 0.001.
  - # Colors:** Set to 10, with step ± 1.
  - Color Assignment:** A table with columns 'Color' and 'Distinguishing Text'.

| Color | Distinguishing Text |
|-------|---------------------|
| Trans | Trans               |
| Green | Green               |
  - Test Data Assignment:** 'Sort by:' dropdown set to 'Name'.
- Right Panel:** Shows 'Displaying image number:' with navigation buttons and a 'Confirm' button.

Two orange callout boxes highlight the '# Colors' field (10) and the 'Distinguishing Text' field (11).

# Using Athena with Your Own Images

## *Define characteristics of your images*

12. Type the text strings identifying the images to be assigned to each color channel.
13. Click 'Test' (i) to load the images and visually confirm (ii) that the images are assigned to the color channels correctly.
14. Use the left & right arrows to move between each zebrafish image file. If the .tif images are multi-page, the menu will allow moving between pages.
15. Click 'Confirm' once you are finished.

### *Info:*

The contrast settings are defined automatically and may not be optimized to visualize some fluorescence signal.

The screenshot displays the Athena software interface for loading and configuring images. The top navigation bar includes 'Settings', 'Loader', 'Protocols', 'Analyzer', and 'Results'. The main interface is divided into several sections:

- Left Sidebar:** Contains loading options: 'Load Multiwell OME Experiment', 'Load Custom' (highlighted), 'Load External', and 'Loading Results'.
- Central Configuration Panel:**
  - Load From Folder:** Path: C:\Data\My Zebrafish Images
  - Data Format:** Pixel size: 3.196 (mic), # Colors: 2
  - Color Assignment:** Trans (Color) -> ch01 (Distinguishing Text); Green (Color) -> ch00 (Distinguishing Text)
  - Test Data Assignment:** Sort by: Name
- Right Preview Area:** Shows two image channels: 'Trans' and 'Green'. A blue information icon is visible next to the 'Green' channel.

Orange callout boxes with numbers 12, 13.i, 13.ii, 14, and 15 are overlaid on the interface to indicate the steps described in the text.

# Using Athena with Your Own Images

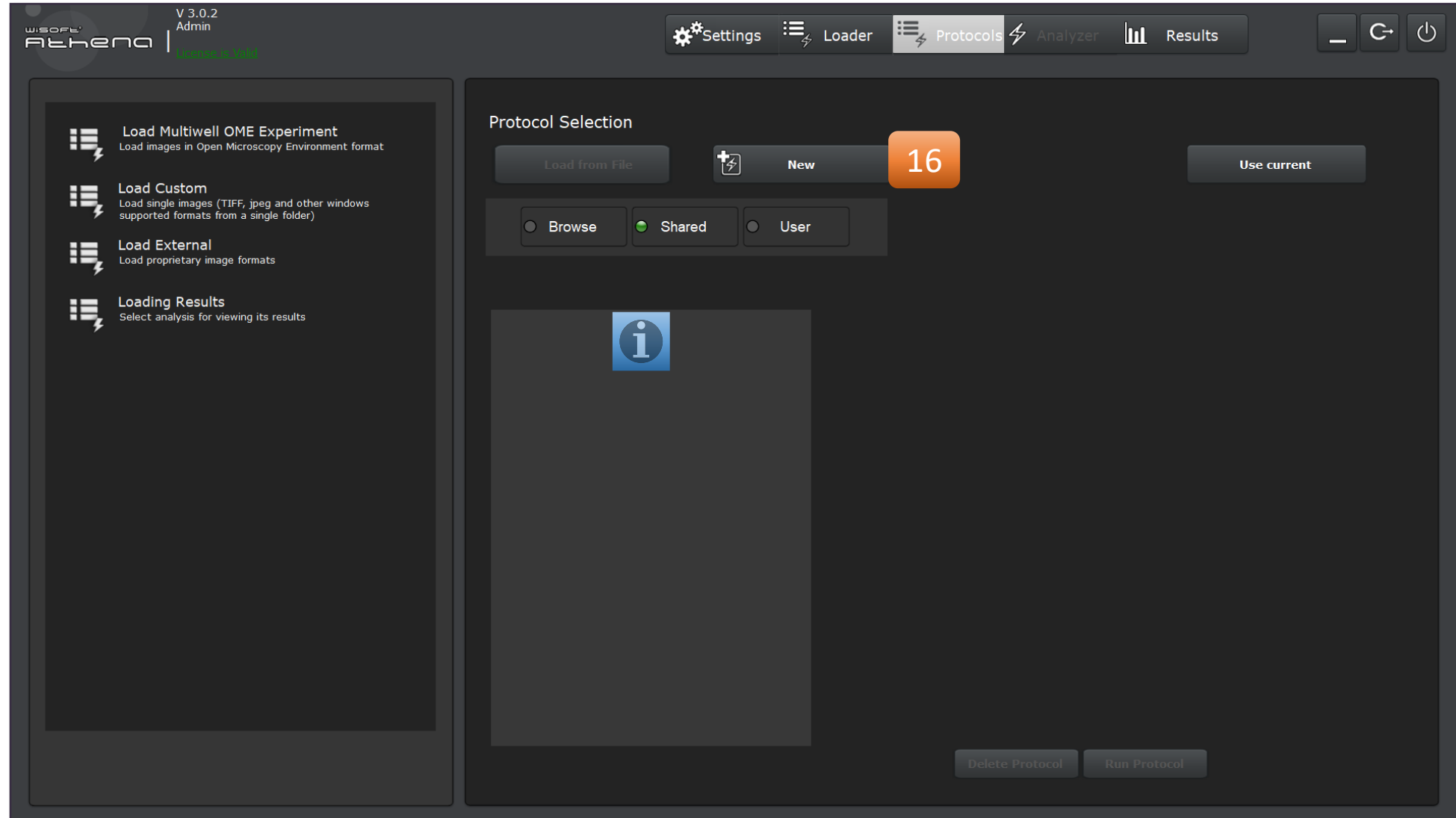
## Protocol menu

16. Click 'New' button to create a new image analysis protocol.

### Info:

Analysis protocols allow for quick, reproducible analysis of images acquired in the same fashion for the same experiment.

They are saved as files that can be loaded from a 'Shared' folder available to all users, a 'User' folder accessible to the user selected in step (3), or can be selected from the hard disk using the 'Browse' option.



# Using Athena with Your Own Images

## Select zebrafish application

17. Select Anatomy to be identified in each color channel.

Light-grey = selected  
Dark-grey = omitted

18. Click 'Next' to advance.

### Info:

The "Other" structure is a flexible anatomical region, that can be manually outlined in brightfield channels or can be identified in a fluorescence channel.

The "Granules" anatomy is intended to be for smaller, point-like structures, such as fluorescently labeled cells.

The "Other" structure can be larger structures, such as fluorescently labeled organs.

WISOFT Athena V 3.0.2 Admin  
License is Valid

Settings Loader Protocols Analyzer Results

Application Selection

Zebrafish  
Fish detection and Quantification

Objective x0

|                                     |                                     |                                     |                                     |                                     |
|-------------------------------------|-------------------------------------|-------------------------------------|-------------------------------------|-------------------------------------|
| Fish                                | Yolk                                | Eye                                 | Fin                                 | Spine                               |
| <input type="checkbox"/>            | <input type="checkbox"/>            | <input type="checkbox"/>            | <input type="checkbox"/>            | <input type="checkbox"/>            |
| <input checked="" type="checkbox"/> | <input checked="" type="checkbox"/> | <input checked="" type="checkbox"/> | <input checked="" type="checkbox"/> | <input checked="" type="checkbox"/> |
| Head                                | Trunk                               | Tail                                | Granules                            | Other                               |
| <input type="checkbox"/>            | <input type="checkbox"/>            | <input type="checkbox"/>            | <input type="checkbox"/>            | <input type="checkbox"/>            |
| <input checked="" type="checkbox"/> | <input checked="" type="checkbox"/> | <input checked="" type="checkbox"/> | <input checked="" type="checkbox"/> | <input checked="" type="checkbox"/> |

Analysis Setup

Next

## Zebrafish Analysis

Automatic quantification of Zebrafish embryo  
for studying of morphological features, fluorescence measurements  
and internal organelle properties

17

18

# Using Athena with Your Own Images

## *Parameter Definition*

Please be patient while the “Processing Masks” flashes, the software is working.

The screenshot displays the Athena software interface for Zebrafish analysis. The top navigation bar includes 'Settings', 'Loader', 'Protocols', 'Analyzer', and 'Results'. The main window is titled 'Analysis Parameters Calibration - Zebrafish' and is divided into several sections:

- Analysis Parameters Calibration - Zebrafish:**
  - Fish:** Minimum Area (100000 mic<sup>2</sup>), Maximum Area (1E+09 mic<sup>2</sup>), Drawing Mode (Auto), and Measure Intensity of (0, 0).
  - Yolk, Eye, Fin, Spine:** Lists of parameters for these features.
  - Defaults:** A button to reset parameters.
  - A1:** A button to select the current analysis.
  - Back/Next:** Navigation buttons.
- Well: A1 F:1 T:1 Z:1:** A vertical list of image thumbnails for different features: Fish [Trans], Yolk, Eye [Trans], Fin [Trans], and Spine.
- Processing Masks:** A large central image of a zebrafish with a grid overlay. A histogram shows the intensity distribution with a threshold line at 336. The histogram data is: Min: 336, Max: 16608, Threshold: [336, 16608]. The current intensity is 10768. A 'Log' checkbox and 'Show Saturation' button are also present.
- Overlay Definition:** A section at the bottom with a 'Well' dropdown and navigation arrows.

# Using Athena with Your Own Images

## Parameter Definition

Identified structures are outlined on the image

19. Anatomy detected in the image are displayed as masks, seen as outlines.
20. Intensity histogram (i) & navigation plate map (ii); minimize them with the arrow-buttons below them.
21. Scroll between different images, each assigned to one well, using the left & right arrow buttons. If T-stack or Z-stack data is loaded, you can also move between different time points or slices.

The screenshot displays the Athena software interface. At the top, the title bar includes 'WISOL Athena V 3.0.2 Admin' and navigation buttons for Settings, Loader, Protocols, Analyzer, and Results. The main window is divided into several panels:

- Left Panel:** 'Analysis Parameters Calibration - Zebrafish'. It contains settings for 'Fish' (Minimum Area: 100000 mic<sup>2</sup>, Maximum Area: 1E+09 mic<sup>2</sup>, Drawing Mode: Auto) and a list of structures: Yolk, Eye, Fin, and Spine. A 'Defaults' button and a well selector (A1) are also present.
- Middle Panel:** 'Well: A1 F:1 T:1 Z:1'. It shows a vertical stack of image thumbnails for 'Fish [Trans]', 'Yolk', 'Eye [Trans]', 'Fin [Trans]', and 'Spine', each with a colored dot and a scroll bar.
- Main Image:** A large grayscale image of a zebrafish embryo with colored outlines (green, purple, yellow) representing detected structures. An orange callout '19' is positioned over the image.
- Bottom Right:** An intensity histogram (labeled '20.i') and a navigation plate map (labeled '20.ii'). The histogram shows a peak with 'Min: 336', 'Max: 16608', and 'Threshold: [336, 16608]'. The plate map is a 12x12 grid with a red dot in the top-left corner.
- Bottom Center:** A scale bar indicating '800µm (72.6%)' and a well selector dropdown set to 'Well'. An orange callout '21' is positioned below the well selector.

# Using Athena with Your Own Images

## *License vs. Trial version*

Identified structures are quantified

19. When highlighting a mask, as the fish here, it becomes selected.

If you are using a **Licensed** version of the software, then quantitative data is made available and is displayed at the bottom of the screen.

The screenshot displays the Athena software interface for image analysis. The main window shows a grayscale image of a zebrafish with various parts segmented and highlighted in different colors. A large orange box with the number '19' is overlaid on the fish's body. The interface includes a top navigation bar with 'Settings', 'Loader', 'Protocols', 'Analyzer', and 'Results' tabs. On the left, there is a 'Analysis Parameters Calibration - Zebrafish' panel with settings for 'Fish', 'Yolk', 'Eye', 'Fin', and 'Spine'. The 'Fish' section has 'Minimum Area' set to 100000 mic<sup>2</sup> and 'Maximum Area' set to 1E+09 mic<sup>2</sup>. Below this is a 'Drawing Mode' dropdown set to 'Auto' and 'Measure Intensity of:' controls. A 'Well: A1 F:1 T:1 Z:1' panel on the left shows a list of segmented parts: Fish [Trans], Yolk, Eye [Trans], Fin [Trans], and Spine. At the bottom right, a histogram shows the intensity distribution of the selected mask, with 'Min: 336', 'Max: 16608', and 'Threshold: [336, 16608]'. A 'Grid' panel shows a 12x12 grid of cells. The bottom status bar displays quantitative data: 'Fish Area: 1.44 mm<sup>2</sup>', 'Count Eye: 2', 'Count Fin: 1', 'Count Head: 1', 'Count Tail: 1', 'Count Yolk: 1', 'Count Spine: 2', 'Count Trunk: 1', 'Eye Area: 4.6e+4 mic<sup>2</sup>', 'Fin Area: 7759.03 mic<sup>2</sup>', and 'Head Area: 0.35 mm<sup>2</sup>'. The bottom left has 'Back' and 'Next' buttons.

# Using Athena with Your Own Images

## *License vs. Trial version*

Identified structures are quantified

19. When highlighting a mask, as the fish here, it becomes selected.

If you are using a **Licensed** version of the software, then quantitative data is made available and is displayed at the bottom of the screen.

Quantitative data is also accessible via the 'Attributes' menu accessed by right-clicking on the image.

More data is accessible on the final Results screen.

The screenshot displays the Athena software interface. On the left, the 'Analysis Parameters Calibration - Zebrafish' panel shows settings for Fish (Minimum Area: 100000 mic<sup>2</sup>, Maximum Area: 1E+09 mic<sup>2</sup>, Drawing Mode: Auto, Measure Intensity of: 0), and other structures like Yolk, Eye, Fin, and Spine. A 'Defaults' button is visible. The main window shows a grayscale image of a zebrafish with a red mask and a blue number '19' overlaid. A context menu is open over the fish, with 'Attributes' selected. A histogram is visible in the bottom right. The bottom status bar shows 'Fish Area: 1.44mm<sup>2</sup>' and a summary of counts and areas for various structures.

**Attributes Window Data:**

- Fish Area: 1.42037 mm<sup>2</sup>
- Fish Axial Ratio: 5.72794
- Fish Long Axis: 1.29659 mm
- Fish Perimeter: 7.87054 mm
- Fish Solidity: 0.75913
- Fish Variance: 6.99e+6
- Count Eye: 2
- Eye Area: 46288.05859 mic<sup>2</sup>
- Count Fin: 1
- Fin Area: 7759.0332 mic<sup>2</sup>
- Count Granules: 9
- Granules Area: 54.59303 mic<sup>2</sup>
- Granules Tot Int: 43235.55469
- Granules Tot Int - Bck: 40428.62891
- Count Head: 1
- Head Area: 0.35368 mm<sup>2</sup>
- Count Spine: 2
- Spine Area: 92750.13281 mic<sup>2</sup>
- Count Tail: 1
- Tail Area: 0.38554 mm<sup>2</sup>
- Count Trunk: 1
- Trunk Area: 0.67482 mm<sup>2</sup>
- Count Yolk: 1
- Yolk Area: 0.218 mm<sup>2</sup>

**Status Bar Summary:**

|               |                                   |                                    |
|---------------|-----------------------------------|------------------------------------|
| Count Eye: 2  | Count Yolk: 1                     | Fish Area: 1.42 mm <sup>2</sup>    |
| Count Fin: 1  | Count Spine: 2                    | Fin Area: 7759.03 mic <sup>2</sup> |
| Count Head: 1 | Count Trunk: 1                    | Count Granules: 9                  |
| Count Tail: 1 | Eye Area: 4.6e+4 mic <sup>2</sup> | Head Area: 0.35 mm <sup>2</sup>    |

# Using Athena with Your Own Images

## License vs. Trial version

Identified structures are quantified

19. When highlighting a mask, as the fish here, it becomes selected.

If you are using a **Trial** version of the software, then no quantitative data is made available.

Email [info@idea-bio.com](mailto:info@idea-bio.com) to purchase a license package and get the data you need.

### Athena Packages

#### Flexible Pay-Per-Use License

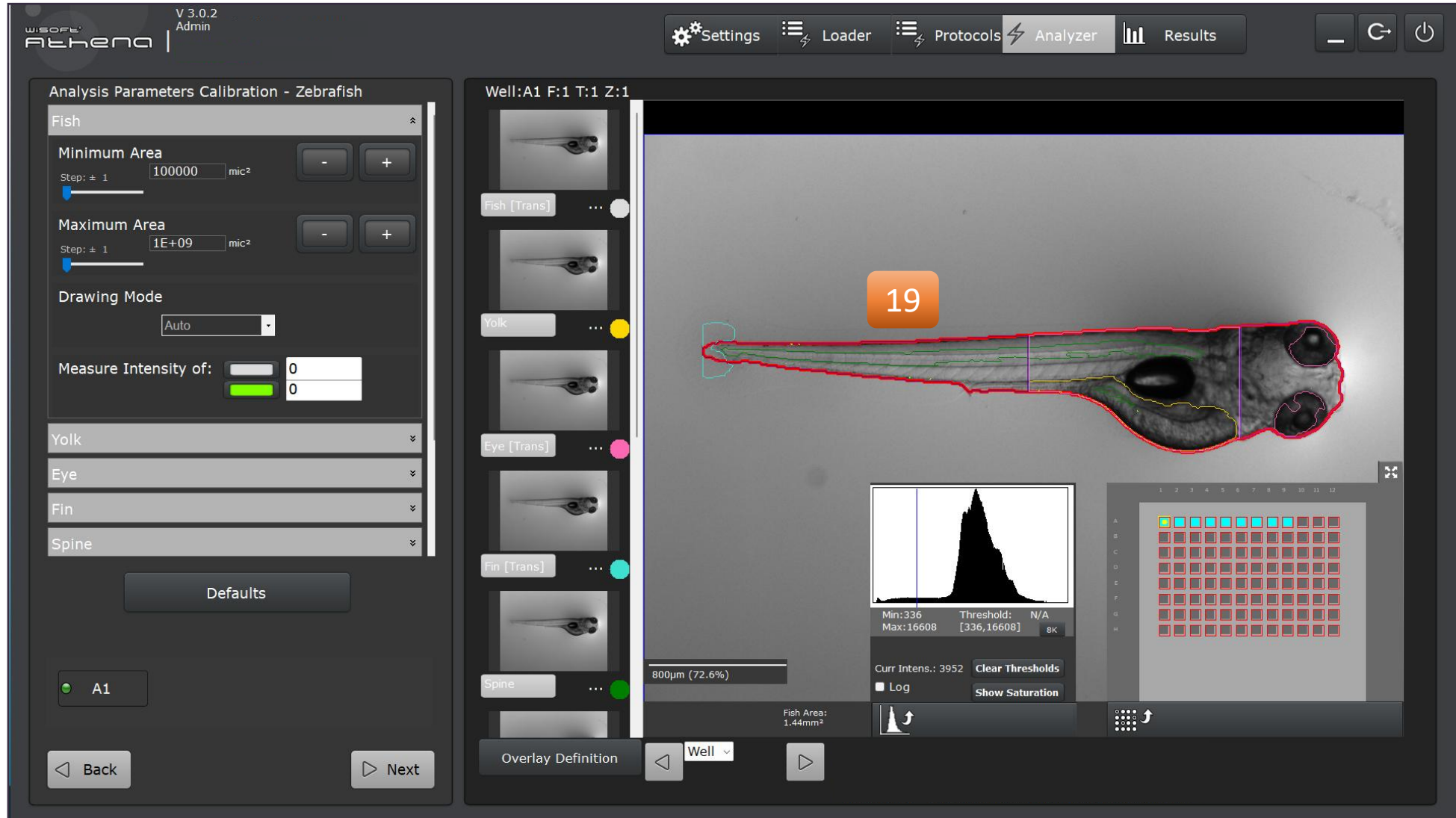
We offer six license packages for different throughput requirements. Choose the best package that fits your needs.

| Number Of Images: |          |       |        |        |            |
|-------------------|----------|-------|--------|--------|------------|
| 500               | 1,000    | 5,000 | 10,000 | 25,000 | 50,000     |
| Starter           | Advanced | Pro   | Expert | Elite  | Enterprise |

**\$150**  
Price per image: \$0.30

[Order Now](#)

\*All purchased packages are valid for 12 months post installation



Click above for more info.



# Using Athena with Sample Images

## *Signal Integration Within Structures*

24. Using the pixel intensity histogram (20.i), drag left-to-right to define the visualized contrast levels.

When all fluorescence objects of interest are visible in white or grey color, then the lower pixel intensity bound can be used for signal integration.

In this example, the minimum value is 949 AU, highlighted with an orange circle.

The screenshot displays the Athena software interface (V 3.0.2 Admin) with the following components:

- Top Navigation:** Settings, Loader, Protocols, Analyzer, Results.
- Left Panel (Analysis Parameters Calibration - Zebrafish):**
  - Granules Smooth: Step: ± 0.1, 5 mic
  - Granules Background Subtraction: Step: ± 0.1, 30 mic
  - Granules Intensity Threshold: Step: ± 1, 2700
  - Granules Maximum Merge Area: Step: ± 1, 50 mic<sup>2</sup>
  - Granules Minimum Area: Step: ± 0.1, 10 mic<sup>2</sup>
  - Granules Maximum Area: Step: ± 1, 200 mic<sup>2</sup>
  - Measure Intensity of: 0
  - Defaults button
  - A1 selection button
  - Back and Next navigation buttons
- Center Panel (Well: A1 F:1 T:1 Z:1):**
  - Thumbnail list: Spine, Head, Trunk, Tail [Trans], Granules.
  - Granules thumbnail is highlighted with a green border.
  - Overlay button and Overlay Definition panel.
- Main Image:** A zebrafish image with segmented regions: Spine (green), Head (purple), Trunk (pink), Tail (purple), and Granules (orange).
- Bottom Right Panel (Histogram):**
  - Min: 0, Max: 875605, Threshold: 0.6%, [949, 7048], 5.5.
  - Curr Intens.: 304, Log checkbox checked, Show Saturation button.
  - Clear Thresholds button.
- Bottom Status:** 800µm (72.6%) scale bar, Tail Area: .39mm<sup>2</sup>.

# Using Athena with Sample Images

## *Signal Integration Within Structures*

24. Insert this value into parameter (24) on the fish or other anatomy to integrate fluorescence intensity above this value

The intensity integration can be included for all anatomical regions selected for analysis in the same fashion.

The screenshot displays the Athena software interface (V 3.0.2 Admin) for analyzing a zebrafish image. The interface is divided into several panels:

- Analysis Parameters Calibration - Zebrafish:** This panel on the left contains settings for the 'Fish' region. It includes:
  - Minimum Area:** Set to 100000 mic<sup>2</sup>.
  - Maximum Area:** Set to 1E+09 mic<sup>2</sup>.
  - Drawing Mode:** Set to Auto.
  - Measure Intensity of:** A color scale from 0 (grey) to 949 (green).
  - Regions:** A list of anatomical regions: Fish, Yolk, Eye, Fin, and Spine.
  - Defaults:** A button to reset parameters.
  - Well:** A dropdown menu currently showing 'A1'.
  - Navigation:** Back and Next buttons.
- Well: A1 F:1 T:1 Z:1:** A vertical strip of image thumbnails for different regions: Fish [Trans], Yolk, Eye [Trans], Fin [Trans], and Spine.
- Main Image:** A large grayscale image of a zebrafish with colored overlays (green, purple, yellow) indicating segmented regions.
- Intensity Histogram:** A small graph showing the distribution of fluorescence intensity. The current threshold is set to 0.6%, with a range of [336, 6608].
- Statistics:** Below the histogram, it shows 'Curr Intens.: 11296' and 'Tail Area: .39mm<sup>2</sup>'.
- Buttons:** 'Clear Thresholds', 'Log', and 'Show Saturation' buttons are visible.

# Using Athena with Sample Images

## Identify Fluorescently-labeled Structures

25. Use the 3 image analysis parameters and the 3 object selection parameters to identify fluorescent spots.

### Image analysis

- Smooth
- Background Subtraction
- Thresholding

### Object Selection

- Merge Area  
(joins touching objects)
- Minimum Area
- Maximum Area

26. Click 'Next' to advance.

### *Note:*

*Increasing the smooth or the background subtraction parameters tends to expand masks to detect larger objects.*

*Increasing the threshold parameter tends to shrink the masks and detecting smaller objects.*

The screenshot displays the Athena software interface for image analysis. The top bar includes navigation tabs: Settings, Loader, Protocols, Analyzer, and Results. The main window is titled 'Analysis Parameters Calibration - Zebrafish' and features a sidebar with parameters for 'Granules' analysis. A '25' callout points to the 'Granules Smooth' parameter, which is set to 5 mic. Other parameters include 'Granules Background Subtraction' (30 mic), 'Granules Intensity Threshold' (2700), 'Granules Maximum Merge Area' (50 mic<sup>2</sup>), 'Granules Minimum Area' (10 mic<sup>2</sup>), and 'Granules Maximum Area' (200 mic<sup>2</sup>). A 'Measure Intensity of:' field is set to 0. The 'Defaults' button is visible below the parameters. The main image area shows a zebrafish with segmented structures: Spine (green), Head (purple), Trunk (pink), Tail [Trans] (purple), and Granules (green). A '26' callout points to the 'Next' button at the bottom. The bottom right corner shows a histogram and a table of statistics: Min: 0, Max: 875605, Threshold: 0.6%, and Curr Intens.: 304. The 'Log' checkbox is checked, and 'Show Saturation' is also visible.

# Using Athena with Sample Images

## *Identify Fluorescently-labeled Structures*

27. Optionally, define populations to identify fish having desirable characteristics.

Use of this feature is described in the [Quick Reference Guide](#), so please refer to instructions there for clarification.

28. Click 'Next' to advance.

The screenshot displays the Athena software interface for image analysis. The top navigation bar includes 'Settings', 'Loader', 'Protocols', 'Analyzer', and 'Results'. The main window is titled 'Populations Definition - Zebrafish' and shows a list of defined populations: 'All', 'Fish [Trans]', 'Yolk', 'Eye [Trans]', 'Fin [Trans]', and 'Spine'. Each population has a corresponding colored dot. The central image shows a zebrafish with these regions segmented in various colors. A scale bar indicates '800µm (72.6%)'. The bottom control bar includes 'Back', 'Next', and 'Well' selection options. Two orange callout boxes with the numbers '27' and '28' are overlaid on the interface, corresponding to the text instructions on the left.

# Using Athena with Sample Images

## *Identify Fluorescently-labeled Structures*

29. The summary page allows for review of the analysis parameters defined on previous screens and setting the name of the analysis run.

It also allows for saving of an analysis protocol to facilitate future analysis.

Use of this feature is described in the [Quick Reference Guide](#), so please refer to instructions there for clarification.

WISOFT Athena V 3.0.2 Admin

Settings Loader Protocols Analyzer Results

Application Summary - Zebrafish

Analysis Parameters

8

Fish:  
Minimum Area: 100000  
Maximum Area: 1E+09  
Drawing Mode: Auto  
Measure Intensity of: G

Yolk:  
Minimum Area: 5000  
Maximum Area: 250000  
Drawing Mode: Auto

Eye:  
Minimum Area: 1500  
Maximum Area: 250000  
Drawing Mode: Auto

Fin:  
Minimum Area: 5000  
Maximum Area: 300000  
Drawing Mode: Auto

Spine:  
Minimum Area: 5000  
Maximum Area: 300000  
Drawing Mode: Auto

Head:  
Minimum Area: 5000  
Maximum Area: 1000000  
Drawing Mode: Auto

Trunk:  
Minimum Area: 5000  
Maximum Area: 1300000  
Drawing Mode: Auto

Tail:

Save Protocol Dataset Name: Zebrafish

Back Start

## Zebrafish Analysis

Automatic quantification of Zebrafish embryo  
for studying of morphological features, fluorescence measurements  
and internal organelle properties

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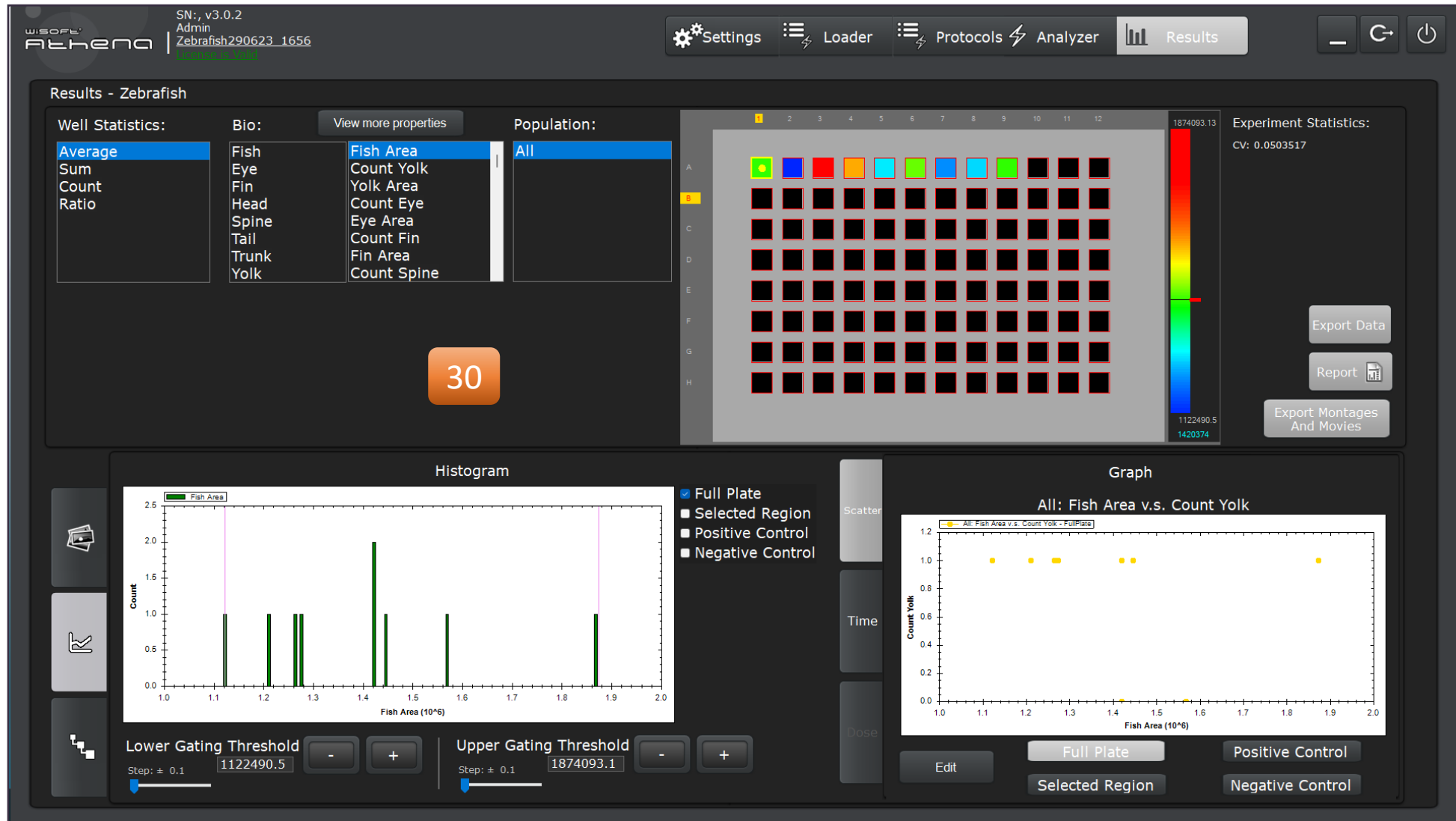
# Using Athena with Sample Images

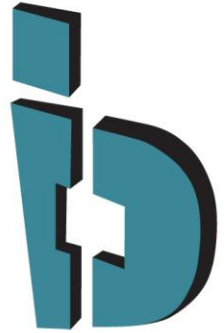
## Identify Fluorescently-labeled Structures

30. The Results page allows for exploration of the data extracted from the zebrafish images (Licensed version only).

It also permits review of the segmentation masks for each image analyzed.

Use of this feature is described in the [Quick Reference Guide](#), so please refer to instructions there for clarification.





# IDEA Bio-Medical

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## Seeing Better



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Need some on-boarding support to get started?

Just email us at [info@idea-bio.com](mailto:info@idea-bio.com) or fill up our [contact form](#).

We'll be sure to contact you soon!